Data supplement for Floris et al., The Link Between Autism and Sex-Related Neuroanatomy, and Associated Cognition and Gene Expression. Am J Psychiatry (doi: 10.1176/appi.ajp.20220194)

Supplemental Information

UK Biobank

Participants, study design and exclusion criteria

UK Biobank (UKB) is a large-scale, population-based, biomedical, epidemiological study comprising around 500,000 predominantly neurotypical (NT) participants. A subset among these has contributed brain imaging data (1). Three dedicated imaging centres are equipped with identical scanner models (3T Siemens Skyra, software VD13) for brain imaging scanning using the standard Siemens 32-channel receive head coil. For details on the brain MRI protocols, see Table 1 in Alfaro-Almagro et al., 2018 (2). In this study, we used the T1-weigthed anatomical MR data from 14,503 individuals (mean age 52.7 years, standard deviation 7.5 years, range 44-80 years), of which 12,949 were used for training, 518 for validation and 1,036 for testing. The image preprocessing pipeline and automated quality control steps are described in detail in (2). We used data as preprocessed already (by our laboratory on behalf of UK Biobank), and as available to all researchers who have been granted access to UKB data. The input data to the deep neural network model was brain-extracted, bias-corrected and registered to MNI152 standard space using affine registration as implemented by FSL-FIRST (3).

ABIDE

Participants, study design and exclusion criteria

We combined the Autism Brain Imaging Data Exchange (ABIDE) repositories I (4) (http://fcon_1000.projects.nitrc.org/indi/abide/abide_I.html) and II (5) (http://fcon_1000.projects.nitrc.org/indi/abide/abide_II.html) for analysis. Sites with same/highly similar imaging acquisition protocols (as outlined and recommended on

the website) were merged across ABIDE I and ABIDE II into one single site (KKI and ABIDEII-KKI; NYU and ABIDEII-NYU; SDSU and ABIDEII-SDSU; UCLA_1 and ABIDEII-UCLA_1). Sites providing less than four individuals (referred to as an individual's imaging data) per sex/diagnostic group were excluded (namely, ABIDEII-BNI_1, ABIDEII-ETH_1, ABIDEII-OILH_2, ABIDEII-SU_2, ABIDEII-TCD_1, ABIDEII-USM_1, ABIDEII-U_MIA_1, CMU, LEUVEN_1, LEUVEN_2, MAX_MUN, OHSU, OLIN, SBL, TRINITY, UCLA_2, UM_2, and USM). Detailed information on imaging acquisition parameters can be found on the above websites. Further, we excluded participants with (clinically non-significant) brain atypicalities (N=2), excessive head motion (N=99) and corrupted image quality (N=3).

For the autism datasets, autism diagnosis was determined by clinician's consensus supported by either one or both 'gold-standard' diagnostic instruments, i.e., an Autism Diagnostic Observation Schedule (ADOS (6)) and/or the Autism Diagnostic Interview-Revised, (ADI-R (7)) in all sites but two (UCD and Stanford sites only used diagnostic cut-offs of ADOS and/or ADI-R for inclusion). This selection process resulted in a total of 1,412 individuals including 115 autistic females, 526 autistic males, 239 NT females, and 532 NT males between 5 and 56 years of age across 15 sites. Individuals were matched for age across and for FIQ within diagnostic groups. For further details, see Table S1.

EU-AIMS LEAP

Participants, study design and exclusion criteria

All participants with autism had an existing clinical diagnosis of autism according to DSM-IV (8), DSM-IV-TR (9), DSM-5 (10) or ICD-10 (11) criteria. Participants underwent comprehensive clinical, cognitive and MRI assessment at one of six

collaborating sites: the Institute of Psychiatry, Psychology and Neuroscience, King's College London (KCL), London, United Kingdom; the Autism Research Centre at the University of Cambridge, Cambridge, United Kingdom; Radboud University Nijmegen Medical Centre, Nijmegen, the Netherlands; University Medical Centre Utrecht, Utrecht, the Netherlands; Central Institute of Mental Health, Mannheim, Germany; and University Campus Bio-Medico, Rome, Italy. Exclusion criteria included the presence of any MRI contraindications (e.g., metal implants, braces, claustrophobia) or failure to give informed written consent to MRI scanning, as well as significant hearing or visual impairments not corrected by glasses or hearing aids. In addition, we excluded participants with missing T1-weighted MRI scans, clinically non-significant brain atypicalities (N= 21), and excessive head motion (N=29). The study was approved by the local ethical committees of participating centers, and written informed consent was obtained from all participants or their legal guardians (for participants<18 years). The final sample comprised 395 autistic individuals (286 males and 109 females), and 286 neurotypical controls (188 males and 98 females) between 6 and 30 years of age. Individuals were matched for age across and for FIQ within diagnostic groups. For details see Table 1 and Figure S1A.

Clinical, cognitive and demographic measures

Intellectual functioning

General intellectual abilities were assessed using the **Wechsler Abbreviated Scales of Intelligence**-Second Edition (WASI-II (12)), or if unavailable, the Wechsler Intelligence Scale for Children-III/IV (WISC-III/IV (13, 14)) for children or Wechsler Adult Intelligence Scale for Adults-III/IV (WAIS-III/IV (15, 16)) for adults.

Standardized estimates of verbal IQ (VIQ), performance IQ (PIQ), and full-scale IQ (FIQ) were derived using IQ norms with mean=100 and SD=15.

Autism-associated, clinical features

The **Autism Diagnostic Observation Schedule** (ADOS-G (6)) was used to measure current, clinically observed core symptoms of autism. Based on the updated algorithm totals (17, 18), we report Calibrated Severity Score (CSS) for 'Social Affect' indexing social-communication difficulties and 'RRBs' indexing restricted and repetitive behaviours. CSS Total serves as an overall indicator of autism severity. The CSS ranges from 1 to 10, with higher scores indicating more substantial autism symptom severity.

The **Autism Diagnostic Interview-Revised** (ADI-R (7)) is a semi-structured caregiver interview completed by parents or caregivers. Algorithm scores were derived from current and historical symptom information for the domains of Reciprocal Social Interaction, Communication, and Restricted, Repetitive and Stereotyped Behaviours and Interests.

The Social Responsiveness Scale, Second Edition (SRS-2 (19)) is a quantitative measure of autistic traits and is composed of 65 items asking about characteristic autistic behaviour over the previous six months. The total raw score is transformed into sex-specific T scores. Parent report was used for all autistic participants and autistic adults additionally completed the self-report form.

The **Repetitive Behaviour Scale**-Revised (RBS-R (20)), composed of 43 items, was used to derive parent-reported total raw scores for restricted and repetitive behaviours, with higher scores indicating a greater level of atypical behaviours.

Sensory processing atypicalities were assessed using the **Short Sensory Profile** (SSP (21)) across 38 items, from which a total raw score was obtained (lower scores indicate more atypicality) that reflect dysfunction across multiple sensory domains.

Adaptive behaviour was assessed with semi-structured parent/caregiver interviews using the **Vineland Adaptive Behaviour Scale**-Second Edition (22). This measures a person's current level of functioning across three domains: communication (expressive, receptive, and written), daily living skills (community, domestic, and personal), and socialization (coping skills, interpersonal relationships, and play and leisure time). For each domain, standard scores were obtained and combined to generate an Adaptive Behavior Composite (ABC) score. Standard scores have a mean of 100 (SD=15), with lower scores indicating greater functional impairment.

Co-occurring conditions (ADHD)

Attention-deficit/hyperactivity disorder (ADHD) symptoms were assessed with the DSM-5 ADHD rating scale, covering both inattention and hyper-activity/impulsivity symptoms based on either self- or parent-report (23). Self-report scores were only used when parent-report scores were unavailable (N=83). A categorical variable was computed based on the DSM-5 criteria (i.e., at least five positive responses in children and six in adults on either or both scales).

Sex-differential cognitive measures

The **Autism Spectrum Quotient** (AQ (24)) is a self- or parent-reported questionnaire that aims to quantify autistic traits. Depending on their age (adults 18-30, adolescents 12-17, children 6-11) and ability level, participants received either an adult (50 items), adolescent (50 items) or child (50 items) version of the test. Before merging the three different versions across all individuals, each version was z-standardized within each diagnostic group and age schedule separately.

The **Empathy Quotient** (EQ) was used to measure a cognitive style described as the drive to identify a person's thoughts and feelings and to respond to these with an appropriate emotion. The **Systemizing Quotient** (SQ) was used to measure a cognitive style characterised by the motivation to predict lawful events (using if-then rules) and observations of input-operation-output relationships and includes good attention to detail. For both, age-appropriate versions were used for children (25), adolescents (26) and adults (27, 28). Before merging the three different versions across all individuals, each version was z-standardized within each diagnostic group and age schedule separately.

The **Reading the Mind in the Eyes** test (29) asks participants to identify complex emotions and mental states based only on the eye region of a face. Depending on their age (adults 18-30, adolescents 12-17, children 6-11) and ability level, participants received either an adult (36 items), adolescent (31 items) or child (28 items) version of the test. Percentage of correct answers was used as the outcome variable. Before merging the three different versions across all individuals, each

version was z-standardized within each diagnostic group and age schedule separately.

Other cognitive measures (selected post-hoc)

Social and monetary reward task

Participants performed a social and monetary incentive delay task within the MRI scanner. Participants were asked to give a speeded response (button press) to a visual target screen flash. For all detail on this task, refer to (30). Behavioural performance on the two tasks was extracted as separate accuracies (i.e., percentage of successful trials) and used as independent variables for our analyses.

Emotional face matching task (Hariri)

Participants completed a well-established face matching task (31) within the MRI scanner, with alternating blocks of faces (showing angry and fearful emotions) and control conditions. In the emotional face condition, a target face has to be matched to one of two probes (identity match) by pressing the left or right button of a response device. Analogously, in the control condition, participants are asked to match a target shape (circle or ellipses) to two test shapes. Behavioural performance on the task was extracted as the accuracy on performing the task (i.e., percentage of successful trials) and used as independent variable for our analyses.

Karolinska Directed Emotional Faces task

The Karolinska Directed Emotional Faces (KDEF) task (32, 33) tests for recognition of basic emotions. Participants were administered an adapted version with 70 trials (34) reduced from the original 140 trials, to reduce assessment time. In each trial,

participants were shown a photograph of a person's face and asked to select which of seven words (happy, sad, angry, surprised, afraid, disgusted, or neutral) best describes the expression displayed. Behavioural performance was defined as the accuracy on the task.

MRI data acquisition

MRI data were acquired on 3T scanners: General Electric MR750 (GE Medical Systems, Milwaukee, WI, USA) at Institute of Psychiatry, Psychology and Neuroscience, King's College London, United Kingdom (KCL); Siemens Magnetom Skyra (Siemens, Erlangen, Germany) at Radboud University Nijmegen Medical Centre, the Netherlands (RUNMC); Siemens Magnetom Verio (Siemens, Erlangen, Germany) at Autism Research Centre at the University of Cambridge, United Kingdom (UCAM); Philips 3T Achieva (Philips Healthcare Systems, Best, The Netherlands) at University Medical Centre Utrecht, the Netherlands (UMCU); GE Medical Systems Signa HDxTt at the Rome University; and Siemens Magnetom Trio (Siemens, Erlangen, Germany) at Central Institute of Mental Health, Mannheim, Germany (CIMH). Procedures were undertaken to optimize the MRI sequences for the best scanner-specific options, and phantoms and travelling heads were employed to assure standardization and quality assurance of the multi-site image-acquisition. Structural images were obtained using a 5.5-minute MPRAGE sequence (TR=2300ms, TE=2.93ms, T1=900ms, voxels size=1.1x1.1x1.2mm, flip angle=9°, matrix size=256x256, FOV=270mm, 176 slices). For further details see Table S3.

Preprocessing

For imaging and data acquisition parameters in UKB see (2), in ABIDE see http://fcon_1000.projects.nitrc.org/indi/abide/ and in LEAP see Table S3. The UKB MRI data have been preprocessed using a standard processing pipeline specified in (2). We used the brain-extracted, bias-corrected and linearly registered (12 degree-of-freedom; to MNI152 standard space) input images to train the convolutional neural network (CNN) / Simple Fully Convolutional Network (SFCN). For ABIDE, LEAP and the ADHD samples, structural T1-weighted images were preprocessed using a standard preprocessing pipeline that included tools from the FMRIB Software Library (FSL version 5.0.6 and 6.0; http://www.fmrib.ox.ac.uk/fsl). After brain extraction using FSL BET (35) and bias-correction, T1-weighted images of each participant were linearly registered to MNI152 standard space using 12-parameter affine transformation. Importantly, brain volumes of all individuals were normalized in this linear registration step.

Sex-Classification

Simple Fully Convolutional Network (SFCN)

We used a previously validated (36) convolutional neural network (CNN) architecture to estimate biological sex based on the preprocessed imaging data. This architecture is based on VGGNet (37) using a fully convolutional structure (38). As previously reported (36) we keep the number of layers to a minimum which significantly reduces the number of parameters to 3 million and consequently also the computational complexity and memory cost. This lightweight deep learning model is referred to as Simple Fully Convolutional Network (SFCN).

The model consists of seven blocks, as shown in Figure S2. Each of the first five blocks contains a 3-by-3-by-3 3D convolutional layer, a batch normalisation layer (39), a max pooling layer and a ReLU activation layer (40). The 1mm-input-resolution 160 × 192 × 160 3D input image (with little or no brain tissue loss) goes through each block sequentially, with its feature map generated and spatial dimension reduced to 5 × 6 × 5 after the fifth block. The sixth block contains a 1 × 1 × 1 3D convolutional layer, a batch normalisation layer and a ReLU activation layer. The seventh block contains an average pooling layer, a dropout layer (only used for training, with 50% random dropout rate) (41), a fully connected layer and a softmax output layer. The channel numbers used in each convolution layer are [32, 64, 64, 64, 64, 64, 2]. Reproducible code can be found at https://github.com/ha-ha-ha-han/UKBiobank_deep_pretrain.

Pre-training in UKB

We pre-trained four CNN models in UKB for sex-classification using the brain-extracted, bias-corrected and linearly registered T1-weighted images as input (Figure S2). The use of such whole brain anatomical data instead of artificial features has been shown in our previous study to have comparable predictive performance across different modalities when using our deep learning model (see Table 5 in (36)). Next, the 14,503 individual brains used for training and validation were randomly divided into four folds for cross-validation, resulting in four CNN models. Each fold contained about one fourth of individuals for testing, and 518 individuals for validation, with remaining individuals used for training. For detailed description of this previously applied training protocol, see (36). All analyses were implemented using

Python and PyTorch (42). Final results revealed a sex prediction accuracy of p=99.5% in UKB (36).

Transfer-learning and validation in ABIDE

Transfer-learning is widely used in medical image analysis given its utility in real world settings where it is expensive or impossible to recollect training data. It does not require the training and test data to be in the same feature space and to have the same distributions. For 2D images, it is common to transfer the models trained with natural images (ImageNet) to medical image datasets (43, 44) to improve the model performance. Similarly, for the 3D MR images, we can use the model pre-trained in a large cohort, and then transfer to smaller samples with different population distribution and different scanner setup, while retaining good predictive performance. Here, we applied transfer learning (45, 46) to transfer the deep learning models trained in UKB to NT individuals in ABIDE. Thus, here, we used the model pre-trained in a large cohort (UKB) and transferred it to a smaller sample (ABIDE) with different distributions (such as age and site) while retaining good predictive performance (44, 45).

Balanced samples across sex, diagnosis and site

To use the data in the most unbiased way for sex-classification in ABIDE (and LEAP/ the ADHD sample further on), we randomly sampled the cohort 100 times for training and validation using the following protocol to get balanced numbers of individuals in each diagnostic (ASC/NT) and sex group (M/F) across all sites: (1) sex matching within each site: we sampled the same number of males and females within NTs,

and the same number of autistic males and females; (2) selecting training and validation set within each sex group: if the selected number (N) of NTs was more than twice the number of autistic individuals, then we used all of the autistic individuals for validation (N ASC), and the same number of NTs for testing (N ASC), while the remaining N of NTs for training (N NT – N ASC); if the selected number of NTs was less than twice the number of autistic individuals, then we used half of the number of NTs for training (N ASC), the remaining half of NTs for validation (N NT – N ASC), and the same number of autistic individuals for validation (see Table S4).

Sex predictions in ABIDE

After this individual sampling, the SFCN models were initialized with the UKB-pre-trained weights (each initialization is shared by 25 models), and next, stochastic gradient descend was used to optimize a loss function of cross-entropy for sex-classification. The learning rate was initially 1e-4 and then decayed by a factor of 10 for every 15 epochs (during one epoch, every training individual is seen once by the model), and the final model was obtained after 50 epochs. The trained model was then used to predict sex for the validation set. For each of the 100 models, the sex-classification accuracy was computed for each sex and diagnostic group (i.e., ASC-M, ASC-F, NT-M, NT-F; for details see below).

Comparing sex prediction accuracies across diagnostic/sex groups

Since we were interested in sex-specific accuracy differences in autism that differ from the reference (NTs), we took the median sex prediction probability (i.e., predictive confidence) of the NT individuals as the sex-classification threshold (i.e.,

the median of the balanced neurotypical sample as baseline reference). Specifically, this means that all 100 individual-level predictions were compared against a sex-classification threshold defined by the median prediction probability across NT males and females. An individual-level prediction probability value greater than the median of NT males and females was classified as ‘male’ vs. a value smaller than the median of the NT males and females was classified as ‘female’. This result was than compared against the real biological sex of each individual and summarized in a Boolean value (True=1 or False=0). The true positive rate (i.e., correctly classified ASC-M) was computed by adding up the true positive values and dividing them by true positive + false negative values, while the true negative rate (i.e., correctly classified ASC-F) was computed by adding up the true negative values and dividing them by true negative + false positive values. Resulting sensitivity and specificity are referred to as sex prediction accuracies within each diagnostic/sex group (ASC-M, ASC-F and NTs). This also implies that the sex prediction accuracy of female and male individuals (used as reference) was the same within the NT cohort. In other words, the true positive rate (in NT-M) and the true negative rate (in NT-F) was the same as the balanced accuracy in NT. Finally, in order to compute differences in the group-level sex prediction accuracies across diagnostic/sex groups, we compared the sex prediction accuracy within each balanced (described above) diagnostic and sex group (i.e., ASC-F vs. ASC-M, ASC-F vs. NT(-F) and ASC-M vs. NT(-M)) with one-sample t tests and associated Cohen’s *d*.

Sex predictions in LEAP and ADHD

After getting the 100 fine-tuned models from ABIDE, we applied the models to every individual in LEAP. The median value of the 100 predictions for every individual was

used to form a final individual-level ensemble-prediction. Again, in order to use the balanced neurotypical sample as baseline reference, the prediction threshold was set to the median prediction probability derived from NT males and NT females. We thus compared the sex prediction accuracy within each diagnostic and sex balanced group, namely, ASC-F vs. ASC-M, ASC-F vs. NT(-F) and ASC-M vs. NT(-M).

The same procedure was next applied to the ADHD sample (see Figure S1D and S1F and Table S4). To ensure comparability, we also computed classification accuracies across autistic individuals from LEAP with and without a co-occurring diagnosis of ADHD.

Region-Aligned Prediction (RAP)

To identify the most predictive and biologically meaningful features that drive our prediction at the brain level, here we employ a novel model interrogation approach – Region-Aligned Prediction (RAP (47)). This method generates spatially resolved estimates of sex prediction accuracy and predicts labels at the brain region level. Specifically, RAP aligns the intermediate feature maps (mp4 in this case, corresponding to the 4th layer of the SFCN) across all individuals in the dataset (including both the training and validation sets), and extracts the feature matrix at one spatial location at a time for all individuals. Note that we do not refer to voxels but spatial locations here, because the node at each location can ‘see’ multiple neighboring voxels in the original space due to the proceeding convolutional operations. These spatial locations are arranged in a 3D lattice just as the original voxels, but with lower spatial resolution. Next, a logistic regression model (an L2-regularized multiple regression model) is trained using the training set feature matrix and applied to the validation set to predict sex. This process is repeated

independently for every spatial location of the intermediate layer, resulting in predictions (i.e., RAP) at every spatial location for every individual. Given that the original CNN was trained 100 times, this generated 100 predictions for every individual. We applied the 100 models in the LEAP dataset, and then generated 100 predictions for every individual, as well as the corresponding RAP maps (spatial prediction for maleness probability). For each spatial location of an individual, we used the median prediction from the 100 RAP maps to generate the ensemble prediction. Here, each spatial location in each person's RAP map consists of probabilistic values between 0 and 1 (where 0 means 'least likely male' and 1 means 'most likely male'). Next, t-maps were computed based on the contrasts NT-M vs. NT-F, ASC-F vs. NT-F and ASC-M vs NT-M. Here, positive t statistics imply a higher male probability in both ASC-M and ASC-F. Thus, thresholding these maps will yield regions with highest male probability and thus most informative for the prediction. The RAP generation steps for LEAP dataset are summarized as follows:

1. Every spatial location of each individual gets 100 predictions for the maleness probability.
2. The 100 predictions are summarized with the median value.
3. The process is repeated for all 681 individuals and every individual receives a median-RAP map.

Associations with autism-associated, clinical features

To assess the clinical relevance of our findings, we followed results up in autistic individuals (males and females separately) within the deeply phenotyped LEAP sample. We ran two general linear models (GLMs) for autistic males and females separately, with core clinical and cognitive measures associated with autism as

independent variables while accounting for age and site. These core clinical and cognitive measures included: ADOS (CSS, CSS-SA, CSS-RRB), ADI-R (reciprocal social interaction, communication, RRB), SRS-2, RBS-R, SSP, Vineland Adaptive Behaviour Composite (ABC) Score and three subscales (communication, daily living skills, socialization). We predicted more autistic symptoms and cognitive difficulties with higher male prediction probability and thus computed one-tailed p-values. All results were FDR-corrected. Results revealed no significant associations.

Associations with sex-differential, cognitive measures

To assess the relationship with sex-differential cognitive features, we followed results up in autistic individuals (males and females separately) within the deeply phenotyped LEAP sample. We ran two GLMs for autistic males and females separately, with sex-differential, cognitive features as independent variables while accounting for age and site. These sex-differential, cognitive features included: the Reading the Mind in the Eyes Test (RMET), Autism Quotient (AQ), Empathy Quotient (EQ), and Systemizing Quotient (SQ). We predicted more male-typical scores (i.e., higher scores on the AQ and SQ, while lower scores on the RMET and EQ) with higher male prediction probability and thus computed one-tailed p-values. All results were FDR-corrected. These analyses yielded a nominally significant relationships between higher male sex prediction values with higher scores on the AQ in ASC-M (t=2.05, p=0.02, q=0.08), while a significant relationship with lower performance on the RMET test in ASC-F (t=-2.36, p=0.01, q=0.04, Figure S3A).

We further tested whether a shift towards maleness was also associated with more sex-differential cognitive measures in neurotypical individuals. We ran two GLMs – one in NT males and one in NT females separately with sex prediction

values as dependent variables and age and site as covariates. We predicted that higher sex prediction probability values towards maleness would be associated with higher AQ and SQ scores, and lower EQ scores or lower accuracy on the RMET. We found no significant associations between greater predictive male probability and any of the cognitive measures in NT.

Cognitive decoding

We further investigated two specific RAP-imaging-t-maps (ASC-F vs. NT-F and ASC-M vs. NT-M) focussing on regions with *positive* t values (as these represented where ASC-M and ASC-F showed higher male prediction probabilities). To explore the cognitive domains implicated in these, we used the Neurosynth Image Decoder (http://neurosynth.org/decode/; accessed on January 23rd 2021) to visualize the top 100 terms most strongly associated with the two RAP-imaging-t-maps.
After excluding anatomical (e.g., fusiform gyrus) and redundant terms (e.g., object and objects), we visualized the remaining terms showing correlations with the imaging maps between r=0.2–0.06.

Results showed that the most common cognitive terms associated with the female RAP-t-maps were primarily related to face perception, visual processing and speech (Figure 2D), whereas in males to motor and reward processing (Figure 2E). Based on these results we tested the association between the RAP-based sex prediction values (extracted from clusters with highest male prediction probabilities) and *a)* cognitive measures associated with motor and reward processes in males (i.e., ADI-R RRB, ADOS-2 CSS-RRB, RBS-R, measures of accuracy on monetary and social reward tasks) and *b)* cognitive measures related to face and communication in females (i.e., emotional face processing tasks [KDEF & Hariri],

ADI-R communication and communicative adaptive functioning). Next, in order to validate the specificity of RAP images relative to behavioural data, we cluster-corrected these two RAP-imaging-maps (Gaussian random field theory, voxel-level Z=2.3, cluster-forming p=0.05) to identify ROIs with highest male prediction probabilities in ASC-M and ASC-F. These were then binarized and used as masks (one male-specific and one female-specific mask) to extract the average RAP-values for each male/female autistic individual. These ROI-specific values were then submitted to two GLMs in autistic males and females separately to test the association between the RAP-based sex prediction values within the clusters with the highest male prediction probabilities and with different cognitive measures related to the cognitive domains identified in the previously described cognitive decoding analyses. Results were FDR-corrected. While we found no relationships in ASC-M, in ASC-F there was a significant association between predicted maleness and lower accuracy on the Karolinska Directed Emotional Faces task (t=-2.6, p=0.01, q=0.02) (Figure S3B).

ADHD sample

To assess specificity of results with regards to autism, we selected an independent sample of individuals with attention-deficit/hyper-activity disorder (ADHD) and NT individuals. For this we combined the publicly available ADHD200 (48) (excluding the NeuroImage site collection) and the local NeuroImage (49) sample from Nijmegen and Utrecht (as the local sample is larger in size than the Nijmegen-NeuroImage sample, which is part of ADHD200). Given that the NeuroImage study is a sibling designed cohort, there were pairs of siblings who were either both control individuals or both individuals with a diagnosis of ADHD. We thus created a custom-code that

randomly excluded one of the two sibling pairs, retaining only one member of the family (in both the control and ADHD category). Acquisition sites that provided less than four individuals (referred to as an individual's imaging data) per sex/diagnostic group were excluded (in this case two sites, namely, University of Pittsburgh and Washington University in St. Louis from ADHD200). We further excluded participants with (clinically non-significant) reported atypicalities in the structural MRI data (N=7) and excessive head motion or corrupted image quality (N=54). This resulted in a sample of 370 males with ADHD, 134 females with ADHD, 246 NT males and 220 NT females across six sites in total.

To account for a significant difference in age *(F*=1.7, *p*<0.001) between males and females with ADHD (mean age females=13.3, mean age males=14.2) and NT males and females (mean age females=12.5, mean age males=12.2), we used both auto-age matching implemented in our code (as described below) and we also manually matched individuals on age (mean age females with ADHD=13, mean age males with ADHD=13.3, mean age NT females=12.8, mean age NT males=12.6; *F*=1.97, *p*=0.12; after excluding 4 females with ADHD, 46 males with ADHD, 13 NT females, 21 NT males). This resulted in a final sample of 324 males with ADHD, 130 females with ADHD, 225 NT males and 208 NT females across six sites in total. Individuals were matched for age across sex/diagnostic groups and for FIQ within diagnostic groups (i.e., ADHD-M matched with ADHD-F; and NT-M matched with NT-F). For details, see Table S2.

Sex prediction analyses in the ADHD sample

We applied the 100 fine-tuned models from ABIDE to the combined ADHD200 - Neuroimage sample the same way as described in the method sections in relation to

the LEAP sample. More specifically, the models were applied to every individual in the ADHD testing dataset. Given the age distribution of the sample, we did not want results to be biased by older males and females with ADHD. Thus, we ran analyses in two samples: a) in the manually age-matched sample (as described above) and b) in a sample that was auto age-matched within the code when creating the sex- and clinically-balanced samples for testing. Figure S1D shows the resulting age distributions for the sex- and clinically-balanced sample when doing manual age-matching and Figure S1F shows the resulting age distributions for the sex- and clinically-balanced sample when doing auto age-matching within the code.

Manual age-matching (before the model application)

Results revealed that females with ADHD (72%) had similar sex prediction accuracies as males with ADHD (73%) (Cohen's *d*=0.1, *p*=0.3), and slightly reduced compared to NT individuals (73%) (*d*=0.24, *p*=0.02), however this did not survive Bonferroni correction (*p*=0.05/3=0.017). Males with ADHD showed no differences in sex prediction accuracy from NT (d=0.05, *p*=0.6) (Figure S4A and Table S5A).

Auto age-matching (within the model application)

Results revealed that females with ADHD (72%) had similar sex prediction accuracies as males with ADHD (71%) (Cohen's *d*=0.14, *p*=0.2) and as NT (73%) (*d*=0.19, *p*=0.05). Males with ADHD showed slightly reduced sex prediction accuracies compared to NT (*d*=0.41, *p*=1.0e-04) (Figure S4B and Table S5B).

Comparison of autistic individuals with and without ADHD

To compare results in the LEAP sample with the ADHD sample results, we also computed classification accuracies across individuals with autism with and without a co-occurring diagnosis of ADHD (ADHD+ vs. ADHD-). Autistic individuals were sub-grouped based on DSM-5 criteria as specified above. This resulted in a sample of 120 autistic males with ADHD, 119 autistic males without ADHD (47 autistic males had missing values on the ADHD measure), 39 autistic females with ADHD and 61 autistic females without ADHD (9 autistic females had missing values on the ADHD measure). Note that 13 NT males and 11 NT females also scored above threshold on the ADHD rating scale, were however not excluded from analyses for consistency reasons. For accuracy comparisons between each diagnostic and sex group, the same number of individuals was sampled for each model, i.e., the smallest available number in a certain group, which was 39 (ASC-F) in this case.

Results revealed that the previously reported pattern was present across both autistic males and females with and without a co-occurring diagnosis of ADHD (Table S6). We found that the sex prediction accuracy in autistic females with ADHD (71%) was on average lower than that in autistic males with ADHD (87%) (Cohen's *d*=1, *p*=1.4-e16) and less than that in NT (80%) (*d*=-0.72, *p*=1.5e-10). For the male individuals, sex prediction accuracy in autistic males with ADHD was on average better than in NT (*d*=0.71, *p*=2.1e-10) (Figure S5A). Sex prediction accuracy in autistic females without ADHD (75%) was on average less than that in autistic males without ADHD (86%) (*d*=-0.73, *p*=9.8-e11) and that in NT individuals (*d*=-0.45,

p=1.9-e05). For the male individuals, performance in autistic males without ADHD was on average better than that in NT individuals (*d*=-0.64, *p*=6.9-e09) (Figure S5B). Autistic males with ADHD did not differ from autistic males without ADHD (*d*=-0.03, *p*=0.79), whereas autistic females with ADHD had slightly lower sex prediction accuracies than autistic females without ADHD (*d*=-0.02, *p*=0.03) (however not surviving Bonferroni correction *p*=0.05/2=0.025) (Figure S5C).

Sensitivity / Control analyses

All method-based control analyses were done in the ABIDE validation sample, while sample-specific control analyses were carried out in the LEAP sample.

Age

Despite the absence of significant age differences between the sex and diagnostic groups, we still also included an auto age-matching algorithm into our model code to confirm results when the four samples were perfectly matched within the prediction models. The resulting age distribution of the sex- and clinically-balanced LEAP sample (N=340) can be seen in Figure S1E**.** Results remained unchanged to those from the original pipeline (see Figure S6A and Table S7A).

Further, our finding of reduced and superior prediction performance in autistic females and males, respectively, is most pronounced in childhood and decreases throughout development to young adulthood. Only in the last age bin of around 20-30, we do not observe classification differences across groups. We carefully reviewed the possibility of a trivial technical explanation: although we have more

adult individuals in the pre-training step in UKB, the classifier depends mostly on the fine-tuning (training) stage where we retrain all learnable parameters (44). Since the individual age distribution in this final fine-tuned dataset (ABIDE) peaks at around 12-years, any accuracy bias originating from the sample distribution should be around the younger rather than older individuals in our case.

Total intracranial volume

All the imaging inputs were controlled for brain volume through the 12 degree-of-freedom linear registration step in the preprocessing pipeline. Still, we double-checked whether there were any significant group differences between NT-F and ASC-F and between NT-M and ASC-M in total intracranial volume that might have influenced observed sex prediction patterns. First, we compared Freesurfer derived total intracranial volume estimates (before linear registration) across the groups. Two t tests between NT-F and ASC-F (t=-0.74, p=0.46) and between NT-M and ASC-M (t=-1.5, p=0.12) revealed no significant group differences. Next, we included an automatic brain-volume-matching algorithm on top of the age-matching algorithm to ensure balanced age and brain volume across sex and diagnostic groups. More specifically, individuals were divided into age bins spanning five years each. Within each age bin, individuals were ranked by their total intracranial volume, and divided into ten sub-groups. Hence, individuals in each sub-group were within the 10% brain volume rankings across the population within each age bin. We then sampled the same number of individuals from each sex-clinical label for each subgroup, which ensured that the final population was balanced across age and volume. Based on this, we compared the prediction differences between each sex and diagnostic group again. Results remained unchanged (Figure S6B and Table S7B).

Model choice

Autism probability as a function of sex prediction probability

Previously, we established that different diagnostic groups give rise to differential sex prediction accuracies. Here, by reversing the model, we confirm that conversely the different accuracies of sex prediction give rise to a stratification along the diagnostic groupings – even though using non-linear models (where compared to linear models x [independent variable] and y [dependent variable] cannot simply be interchanged). To do this, we investigated whether the probability of having autism was related to the sex predictions (50). We created eight probability bins in steps of 0.125 and determined the sample probability in each bin by dividing the total number of ASC-F (ASC-M) by the total number of females (males) in each bin and ran Spearman's correlations between the sample probability and the predictive sex probability. Next, we subdivided autistic individuals into a) correctly classified females (P<0.5) / males (P>0.5) and b) misclassified females (P>0.5) / males (P<0.5). Within each sex, we compared the proportions of having autism as a function of being correctly or incorrectly classified.

In ASC-F the sample probability of autism increased with increasing predictive probabilities for being classified as male (*rho*=-0.89, p<0.01), while there was no such relationship in ASC-M (*rho*=0.6, p=0.1; Figure S6C, left). ASC-F misclassified as male were significantly more likely to have an autism diagnosis than autistic females correctly classified as female (P=.82 vs. P=.52; χ^2=3.96, p=0.02). There was no such relationship in ASC-M (P=.62 vs. P=0.56; χ^2=1.59, p=0.1).

Principal component analysis with logistic regression

To check model-dependence of our results in ABIDE, we applied an alternative, but simpler method. Following the same train/validation pattern as we applied within the CNN method, we ran the following principal component analysis (PCA) based experiments 100 times: we used 75 principal components explaining most of the variance of the imaging data, and subsequently trained a logistic regression classifier for sex classification. Results showed the same patterns as we observed before: reduced sex prediction accuracy in autistic females compared to autistic males (Cohen's *d*=1.42, *p*=2.8e-25) and compared to NT (*d*=1.39, *p*=8.4e-25), while increased sex prediction accuracy in autistic males compared to NT (*d*=0.71, *p*=3.6e-10), however with significantly reduced prediction performance (Figure S6C, right, and Table S7C). In this regard, we want to point out that PCA combined with logistic regression only takes into account features that contain very limited information (via PCA decomposition), which often reflects the global features in brains. Thus, it is not surprising that the sex prediction accuracy drops considerably compared to the classification results based on our CNN, model which learned non-linear features and classifiers.

RAP validation (mask out salient regions)

To verify the RAP method in ABIDE, we decided to compare sex prediction results when only using the most salient brain regions versus excluding the most salient brain regions in the RAP maps. For this, we generated two masks: one ASC-sensitive and another ASC-insensitive mask. To obtain the ASC-sensitive mask, we

thresholded the two RAP t-maps (map-A: ASC-F vs. NT-F; map-B: ASC-M vs. NT-M) to retain only the 25% highest values within each. Next, we combined the two thresholded maps using logical union (map-C = map-A ∪ map-B), and smoothed and dilated (51) the resulting map-C. The ASC-insensitive mask was the difference between the MNI mask and the ASC-sensitive mask. Both resulting masks contained about 50% of voxels within the MNI brain mask and were thus comparable in size. Next, the RAP method was re-run twice, once within the ASC-sensitive mask and once within the ASC-insensitive mask and prediction accuracies were computed as specified earlier.

Results revealed that the previously observed sex prediction accuracy differences were more pronounced when applying the ASC-sensitive than the ASC-insensitive mask (Table S7D). Specifically, using the ASC-sensitive RAP mask, sex prediction accuracies were significantly lower in autistic females compared to autistic males (Cohen's *d*=1.71, *p*=3.8-e16) and to NT (*d*=1.45, *p*=1.6e-13). Autistic males also showed reduced sex prediction performance compared to NT individuals (*d*=1.09, *p*=6.0e-10) (Figure S6D, left). Using the ASC-insensitive mask, similar patterns albeit with smaller effect sizes emerged (ASC-F vs. ASC-M: *d*=0.63, *p*=5.6e-05; ASC-F vs. NT: *d*=0.33, *p*=2.6e-02; ASC-M vs. NT: *d*=0.55, *p*=3.9e-04; Figure S6D, right).

Comparison of variance in sex prediction accuracies across groups

We compared the ratios of variances of sex prediction accuracies across the groups using the var.test function in R. NT-M had significantly higher variance across

prediction accuracies compared to NT-F (F=0.15, p<0.001). There was also a sex differences in variance in autistic individuals, however with ASC-M having lower variance compared to ASC-F (F=0.4, p<0.001). ASC-F had significantly higher prediction variance compared to NT-F (F=2.22, p<0.001), while there was no significant difference between ASC-M and NT-M (F=0.82, p=0.12).

Previous research shows that the male population displays higher intrinsic variability than females (52). Here we also see higher prediction variability across males than across females. We exclude however the possibility that sex prediction accuracy differences are driven by variance differences: if sample variance was the same across NT-M and NT-F, we would expect a) improved classification accuracies for both NT-M and NT-F (as our CNN was trained to predict maleness), b) even worse classification accuracies for autistic females. Thus, homogeneity of NT-sample variance would even further accentuate our results in autistic females, however, strikingly, our findings of reduced classification accuracies in autistic females hold even at the background of differences in NT-variance.

Comparison of autistic individuals with high and low prediction accuracies

We applied a quartile split on the prediction probability values in autistic males and females to subdivide them into highly misclassified (lower 25% - ‘ASC-M/F low’) and highly correctly classified (upper 25% - ‘ASC-M/F high’) individuals. These two classes of autistic males/females were compared by computing t-maps (ASC-M/F low vs. ASC-M/F high) based on their RAP maps. When comparing the top 25% with

the bottom 25% of autistic males and females on their RAP contrast maps, the most differentiating regions between highly misclassified and highly correctly classified autistic females were in cerebellum, left thalamus, superior frontal gyrus and frontal medial cortex in grey matter, and left retrolenticular part of internal capsule, posterior thalamic radiation, right anterior corona radiata and the splenium of corpus callosum in white matter (Figure S7A). The most differentiating regions between highly misclassified and highly correctly classified autistic males were in bilateral superior parietal lobule, bilateral supplementary motor cortex, precuneus, superior lateral occipital cortex, paracingulate gyrus, frontal medial cortex and frontal pole in grey matter, and bilateral superior cerebellar peduncle and right superior longitudinal fasciculus in white matter (Figure S7B).

Gene expression decoding

We next wanted to address the question of what the potential sources of masculinization were by tapping into the underlying genomic mechanisms.
First, RAP-imaging-t-maps were uploaded on Neurovault (ASC-F vs. NT-F and ASC-M vs. NT-M and NT-F vs. NT-M as reference; see (https://identifiers.org/neurovault.collection:9354). We then used the gene expression dataset from the Allen Human Brain Gene Expression atlas (AHBA) which includes samples from six post-mortem brains (3 Caucasian, 2 African, 1 Hispanic; 1 female) ages 24-57 years. This limited sample size and large variability across age, sex and ethnicity can impact the transcriptional patterns. Previous studies addressing this have shown that results were not driven by one single donor and generalizable beyond the donor brains in the AHBA after rigorously testing the effect of donor selection by

running leave-one-donor-out analyses (53) and using Bayesian random effects analysis which fitted a hierarchical Bayesian regression model using Markov Chain Monte Carlo sampling (54). We employed the gene expression decoding functionality (54) integrated in Neurosynth (55) (https://neurosynth.org) and NeuroVault (56) (https://neurovault.org). This performs in detail the following steps: it first uses the Allen Brain Atlas REST API to download the gene expression data, extracts the MNI coordinates for each sampling site, draws a spherical ROI (4mm) and extracts the average values of the statistical maps within each ROI. This way, the resulting vector of values can next be correlated with the normalized gene expression values to see how similar they are. Here, an approximate random effects analysis calculates the slope of best linear fit for each donor (individually fitted regression lines) and performs a one-sample t test on those estimates to test how consistent the relations between the gene expression and evaluated map values are and identifies genes whose spatial expression patterns are consistently (i.e., across the six donor brains) highly similar to the evaluated maps. Being a similarity analysis, the decoding analysis has two directions. In one direction, we obtain genes with positive t statistics with genes with high expression in areas where there are strong positive values in the RAP-imaging t-maps (i.e., higher male probability), or vice versa, genes with hardly any expression in areas with very low values in the RAP-imaging t-maps (i.e., higher female probability). On the other hand, the decoding analysis also provides genes with negative t statistics – these are genes with high expression in areas where there are strong negative values in the RAP-imaging t-maps (i.e., higher female probability), or vice versa, genes with hardly any expression in areas with strong positive values in the RAP-imaging t-maps (i.e., higher male probability). In our analyses we are interested in the list of genes obtained from the decoding analysis which gives us

genes that have strong positive values in the RAP-imaging t-maps (i.e., higher male probability) and show high gene expression values in those areas (FDR $p<0.05$). For this reason, we only retained genes from this resulting list with positive t statistics and corrected for multiple comparisons (FDR $p<0.05$).

Gene classes in enrichment analyses

After obtaining this list of genes that were highly expressed in spatial patterns throughout the brain and similar to the two sex-differentiation RAP maps, we went on to test the overlap (i.e., enrichment) of our gene lists with a set of relevant classes of genes. We opted for three different gene classes: **1) autism-associated genes** to investigate whether the genetic likelihood for autism overlaps with the male neurophenotype; **2)** genes acting in **prenatal** development as events during the embryonic period have long-lasting effects on both sexual differentiation and susceptibility for neurodevelopmental conditions (prenatal cell types (57)); and **3) sex-differentially expressed genes** in prenatal development and genes **differentially regulated by prenatal sex steroids**.

Autism associated genes

Structural genetic variants

Structural genetic variants included common genetic variants and de novo mutations. Common genetic risk variants were obtained from a large-scale genome-wide association study by Grove et al. (58). SNP based P-values were converted to gene-based P-values using a hg19 genome build using MAGMA (59) which accounts for linkage disequilibrium between SNPs when calculating gene-based P-

values. The identified list of genes was subsequently Bonferroni corrected (ASC commonRV). Genes harbouring rare, de novo variants associated with autism were identified by Satterstrom et al. (60) by whole-exome sequencing of 35,584 samples (11,986 with autism). A total of 102 rare, de novo protein truncating genes associated with autism (FDR adjusted *P*-value < 0.1) were identified (ASC dnPTV). Parikshak et al. (61) identified fetal gene co-expression modules associated with autism genes that have highly similar expression patterns during cortical development (61). They used RNA sequencing on gene expression data from BrainSpan whole-genome transcriptomic data and ran weighted gene co-expression network analysis. Among the five co-expression modules associated with different forms of autism risk, we selected modules 2 and 3 as transcriptional regulators were enriched for rare genetic variants in early fetal development (M2 and M3).

Transcriptionally dysregulated genes

To capture genetic downstream effects, we selected a range of transcriptionally dysregulated genes in autism. Gandal et al. (62) identified differentially expressed genes from an analysis of RNA sequencing data from the PsychENCODE Consortium, using autism post-mortem frontal and temporal cortex tissue (FDR adjusted *P*-value < 0.05) (865 downregulated: ASC DE Downreg, 746 upregulated: ASC DE Upreg) (62).

Parikshak et al. (63) performed rRNA-depleted RNA sequencing using post-mortem frontal and temporal cortical tissue samples from 48 autistic individuals and 49 control individuals. Doing weighted gene co-expression network analysis, they identified transcriptionally dysregulated co-expression modules in autism. Among these, the downregulated modules were enriched in synaptic function and neuronal

genes, whereas upregulated modules were enriched in genes associated with inflammatory pathways and glial functions (764 downregulated: ASC CTX Downreg CoExpMods, 1111 upregulated: ASC CTX Upreg CoExpMods) (63).

Performing single-nucleus RNA sequencing analysis of cortical tissue of individuals with autism, Velmeshev et al. (64) analysed the transcriptomes of single brain cells. They generated 104,559 single-nuclei gene expression profiles (52,556 from control individuals and 52,003 from autistic individuals) and performed unbiased clustering of nuclear profiles and annotated clusters based on expression of known cell type markers. This way they identified genes differentially expressed in autism in specific cell types (ASC Excitatory, ASC Inhibitory, ASC Microglia, ASC Oligodendrocyte, ASC Astrocyte, ASC Endothelial).

Prenatal cell types

This gene list was based on the study by Polioudakis et al. (57) who performed RNA sequencing on 40,000 cells and created a single-cell gene expression atlas of the developing, mid-gestation (gestation weeks 17 to 18) human neocortex. This resulted in the identification of 16 transcriptionally distinct cell groups which clustered by known major biological cell types at this stage of development, including: endothelia (E), excitatory neurons (EN; migrating excitatory, maturing excitatory, maturing excitatory upper enriched, excitatory deep layer 1, excitatory deep layer 2), interneurons (IN; interneuron MGE and interneuron CGE), intermediate progenitors (IM), microglia (M), cycling progenitors (MN; cycling progenitor S-phase and cycling progenitors G2M phase), oligodendrocyte precursors (OPC), pericyte (P), and radial glia (RG; ventricular radial glia and outer radial glia).

Sex-differentially expressed genes

We further surveyed sex-differentially, prenatally expressed genes. For this, we used the same sex-differential gene array expression data from prenatal samples as Werling et al. (65). This was based on a study by Kang et al. (66) who analyzed exon array data from individuals between 16 and 22 post-conception weeks from frontal, temporal and parietal cortex. For genes differentially regulated by sex hormones, we used gene lists from two studies: 1) Quartier et al. (67) conducted an RNA sequencing study in which embryonic neural stem cells were treated with 100nM of dihydrotestosterone (DHT), a potent, non-aromatizable androgen. We used the same list of genes differentially expressed by DHT as Lombardo et al. (68, 69). 2) Willsey et al. (70) conducted RNA sequencing and identified differentially expressed genes in Xenopus whole brain following 17-β-estradiol treatment. We used the list of genes differentially expressed by estrogen.

Enrichment analyses

Analyses examining enrichment were done with a custom code written by MVL (https://github.com/mvlombardo/utils/blob/master/genelistOverlap.R) computing the enrichment odds ratios and hypergeometric p-values for all enrichment hypergeometric tests based on the sum(dhyper) function in R. The background set size for enrichment analyses was set to the number protein decoding genes considered in the Neurosynth Gene Expression Decoding analyses (i.e., 20,787). To avoid biasing our findings towards genes expressed in brain, we conducted another enrichment analysis using a more conservative list of 16,906 background genes based on real estimates of genes expressed in cortical tissue (71) (see Figure S8),

as also done in prior work (72, 73). Only comparison with FDR $p<0.05$ were interpreted further.

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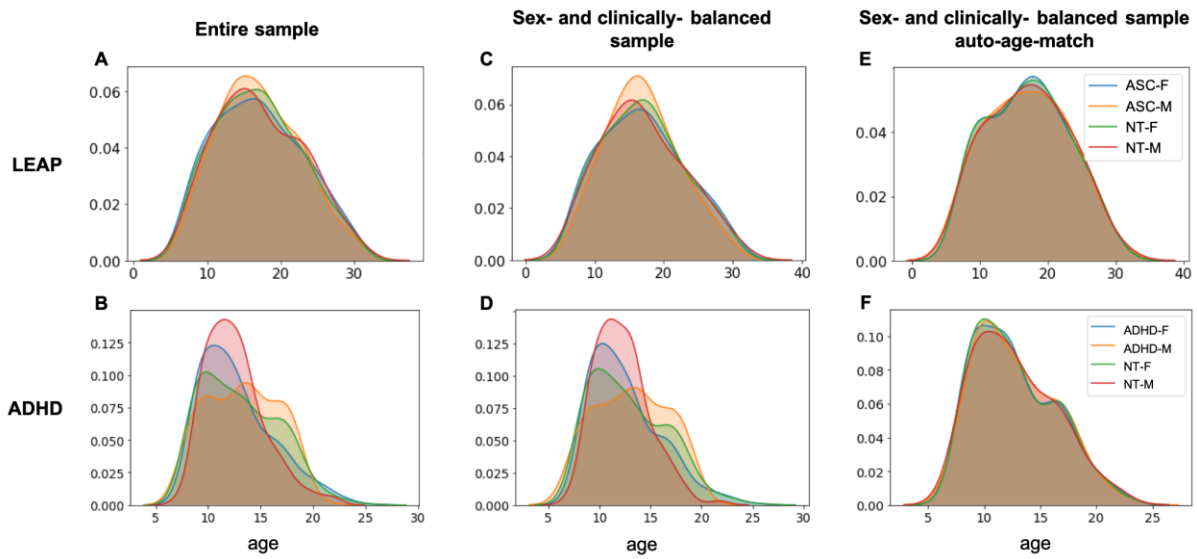
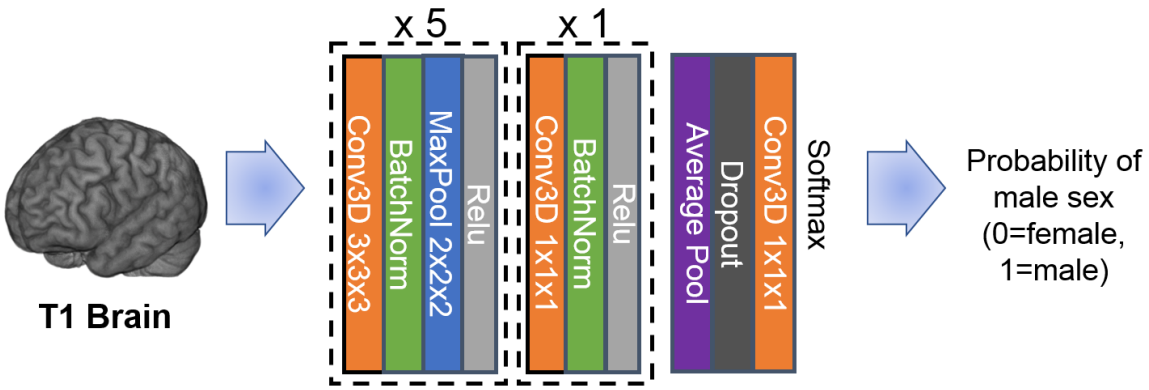
Supplemental Figures

FIGURE S1. Age distributions per sample and model step[a]



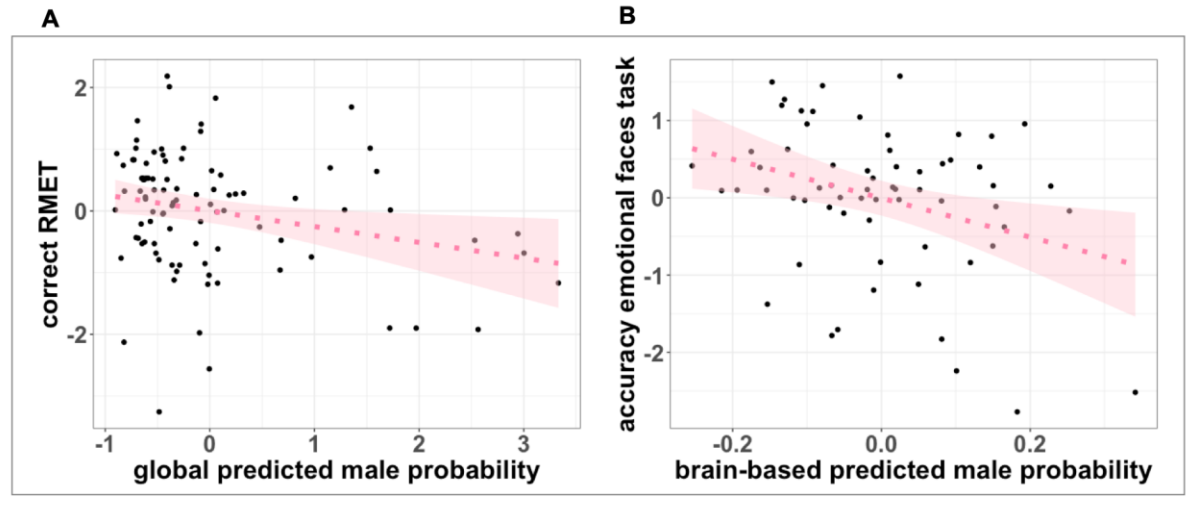
[a] **A**) The age range in the entire LEAP sample comprised 6 to 30 years of age. For details see Table 1**. B**) The age range in the entire ADHD sample comprised 7 to 29 years of age. For details see Table S2. **C**) Age distribution in the sex- and clinically-balanced LEAP sample. **D**) Age distribution in the sex- and clinically-balanced ADHD sample when applying age-matching *before* the model application. **E**) Age distribution in the sex- and clinically-balanced LEAP sample when applying age-matching *within* the model application. **F**) Age distribution in the sex- and clinically-balanced ADHD sample when applying age-matching *within* the model application. As described in the main manuscript, 'sex- and clinically-balanced' refers to randomly sampling the cohort 100 times for training and validation to get balanced, unbiased numbers of individuals in each diagnostic (ASC/ADHD/NT) and sex group (M/F) across all sites. For further details, see Table S4. **Abbreviations**: ASC-F=autistic females, ASC-M=autistic males, NT-F= neurotypical females, NT-M= neurotypical males, ADHD= attention-deficit/hyperactivity disorder, ADHD-F= females with ADHD, ADHD-M=males with ADHD.

FIGURE S2. Simple Fully Convolutional Network[a]



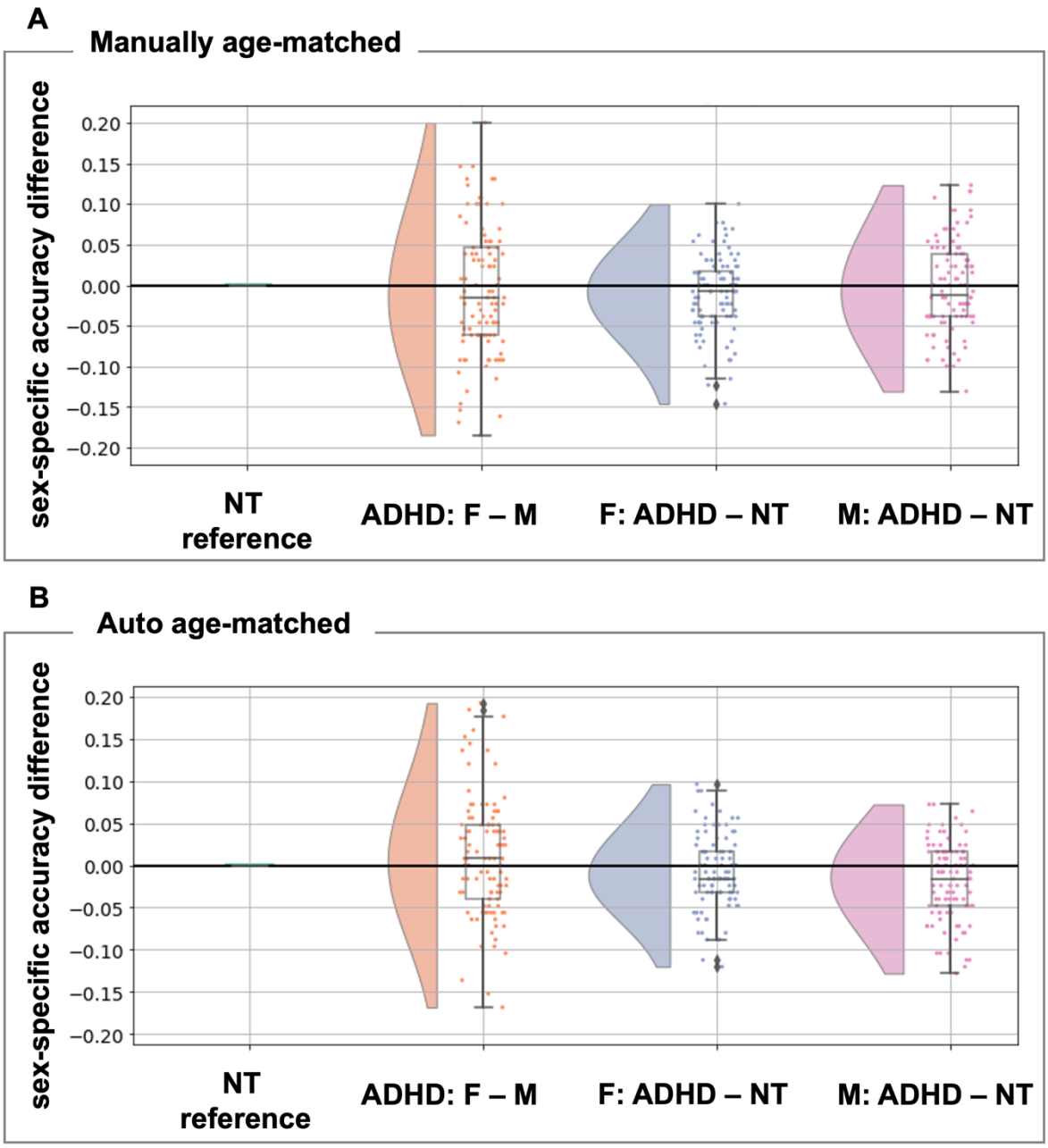
[a] The deep neural network architecture used in this work is a simple fully convolutional neural network (SFCN). The network takes linearly registered MRI brain 3D images as input. It consists of five Conv3D[3x3x3]-BatchNorm-MaxPool[2x2x2]-ReLU building blocks, and one Conv3D[1x1x1]-BatchNorm-ReLU building block, and then the average pooling and the final fully connected (FC) layer (effectively implemented as Cov3D[1x1x1]). The FC-layer outputs are two values, converted through a softmax layer to create the final outputs representing the 'probability' of sex prediction (i.e., predictive confidence).

FIGURE S3. Cognitive associations[a]



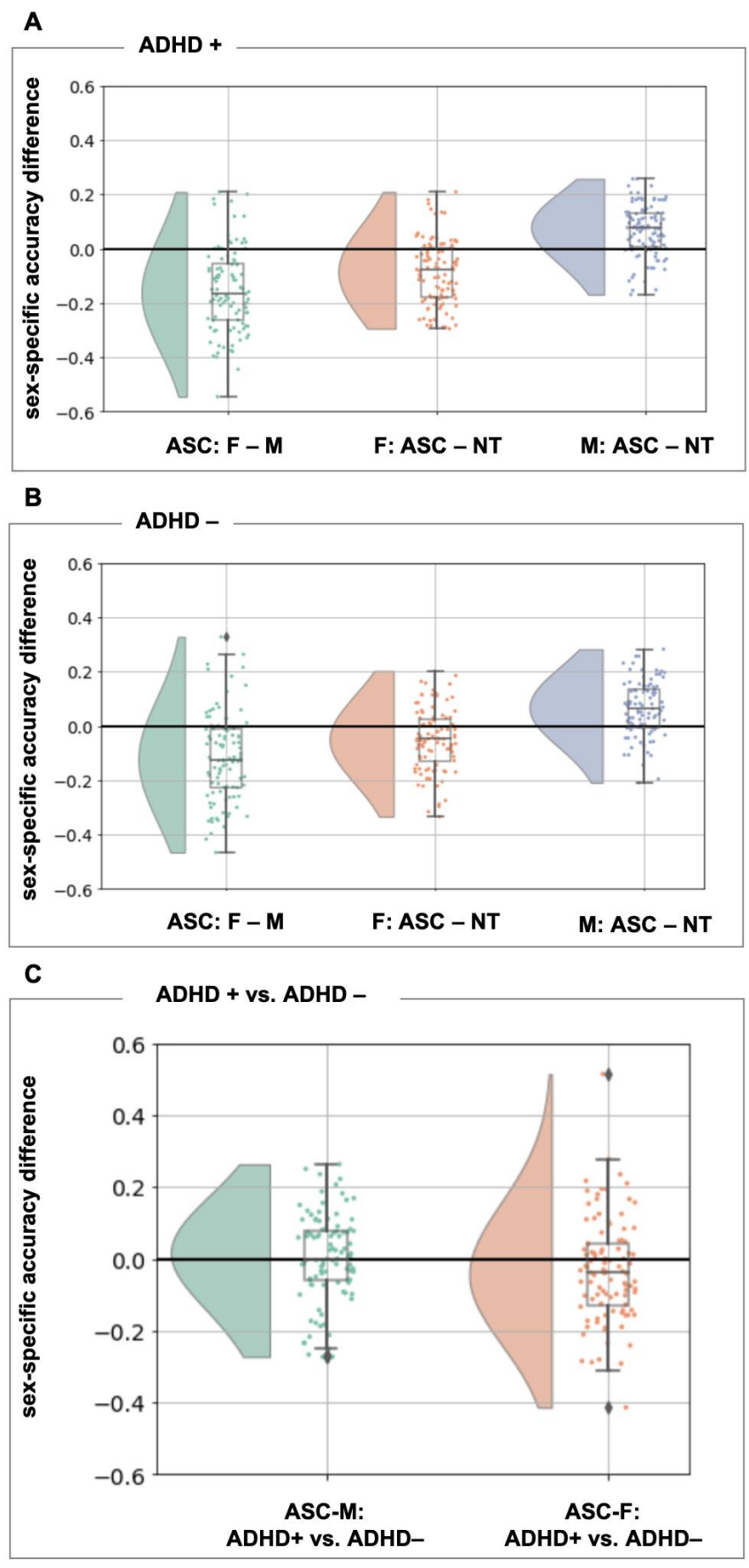
[a] Associations between predicted male probability (i.e., predictive confidence) and sex-differential cognitive features. **A**) Association between the global sex prediction probabilities and correct performance on the Reading the Mind in the Eyes task (RMET) in autistic females. **B**) Association between values extracted from the female RAP-imaging-t-maps (brain-based sex prediction values) and emotional face recognition accuracy (KDEF) in autistic values. Plotted values are z-standardized.

FIGURE S4. ADHD sample prediction accuracies[a]



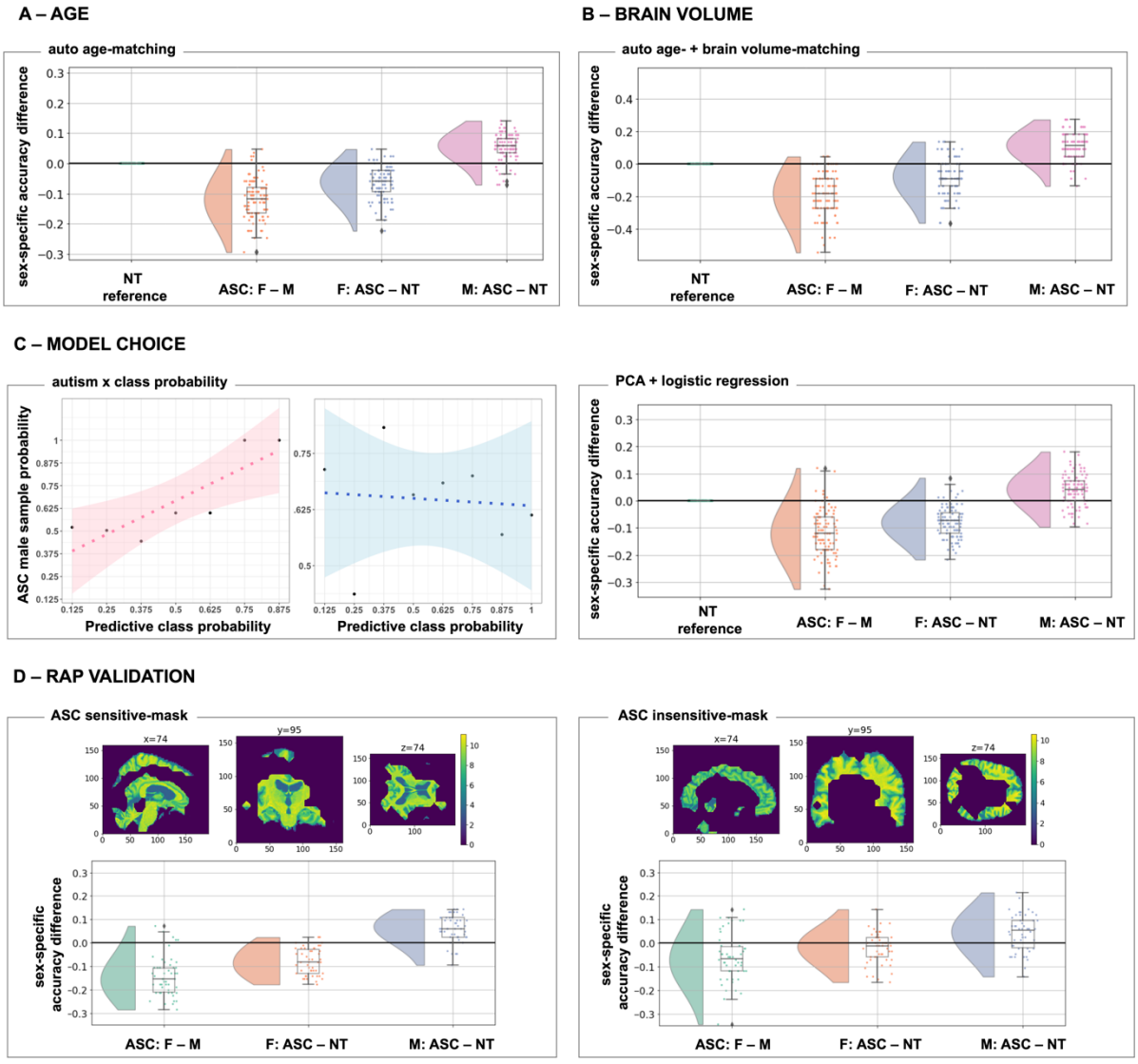
[a] Comparison of sex prediction model performance across diagnostic and sex groups. We compare sex prediction accuracy differences between males and females with ADHD (ADHD: F – M), females with ADHD and neurotypical females (F: ADHD – NT) and males with ADHD and neurotypical males (M: ADHD – NT) in both the clinically- and sex-balanced sample matched for age before model application (**A**) and in the clinically- and sex-balanced sample matched for age within the model application (**B**). Each dot represents one model out of 100 models in total. Negative values mean the model performs worse in the first sex/diagnostic group. Abbreviations: NT=neurotypical, ADHD=attention-deficit/hyperactivity disorder, F=females, M=males.

FIGURE S5. Comparison of autistic individuals with and without ADHD[a]



[a] Comparison of sex prediction model performance across diagnostic and sex groups. We compare sex prediction accuracy differences between autistic males and autistic females (ASC: F – M), autistic females and neurotypical females (F: ASC – NT) and autistic males and neurotypical males (M: ASC – NT) in LEAP including either autistic individuals with co-occurring ADHD (**A**) or autistic individuals without co-occurring ADHD (**B**). In **C**) we compare sex prediction model performance between autistic males with and without ADHD and autistic females with and without ADHD. Each dot represents one model out of 100 models in total. Negative values mean the model performs worse in the first sex/diagnostic group. Abbreviations: ADHD+=autistic individuals with co-occurring ADHD, ADHD– = autistic individuals without co-occurring ADHD, NT=neurotypical, ASC=autism, ADHD=attention-deficit/hyperactivity disorder, F=females, M=males.

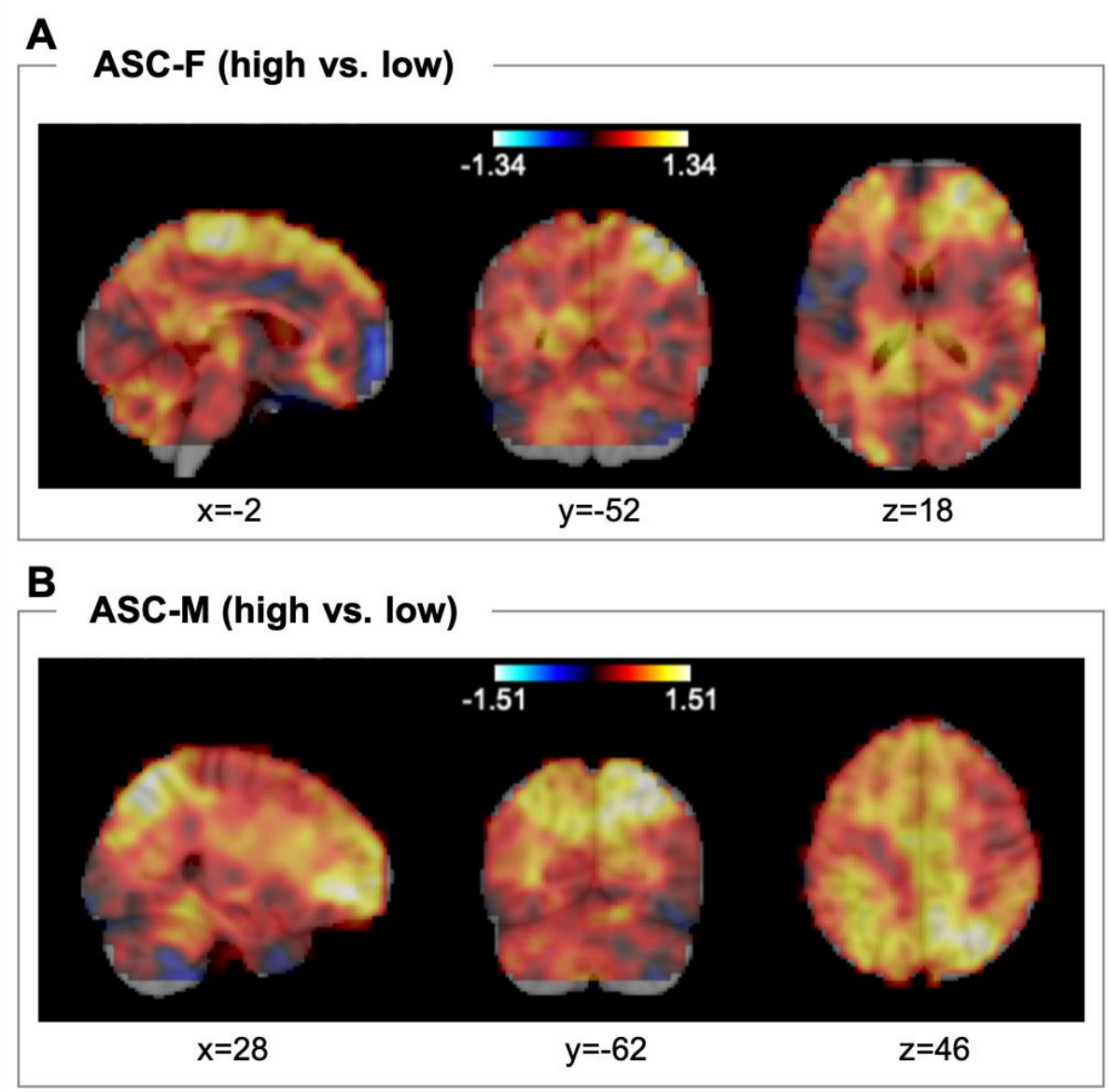
FIGURE S6. Sensitivity and control analyses[a]



[a] Overview of all sensitivity and control analyses. **A**) AGE. We included an auto age-matching algorithm into our model code to confirm results when the four samples were perfectly matched for age within the sex prediction models. We plot the sex prediction model performance across diagnostic and sex groups comparing sex prediction accuracy differences between autistic males and autistic females (ASC: F – M), autistic females and neurotypical females (F: ASC – NT) and autistic males and neurotypical males (M: ASC – NT) in LEAP. Each dot represents one model out of 100 models in total. Negative values mean the model performs worse in the first sex/diagnostic group. **B**) TOTAL INTRACRANIAL VOLUME. We extended the auto-age-matching algorithm to also match individuals on total intracranial volume. **C**) MODEL CHOICE. Left: Association between autism likelihood (sample probability) and predictive sex class probabilities. There is an association in autistic females (pink), however not in males (blue). Right: Principal component analysis combined with logistic regression: we used 75 principal components explaining most of the variance of the imaging data, and subsequently trained a logistic regression classifier for sex classification. **D**) RAP VALIDATION. We validated the RAP method in ABIDE

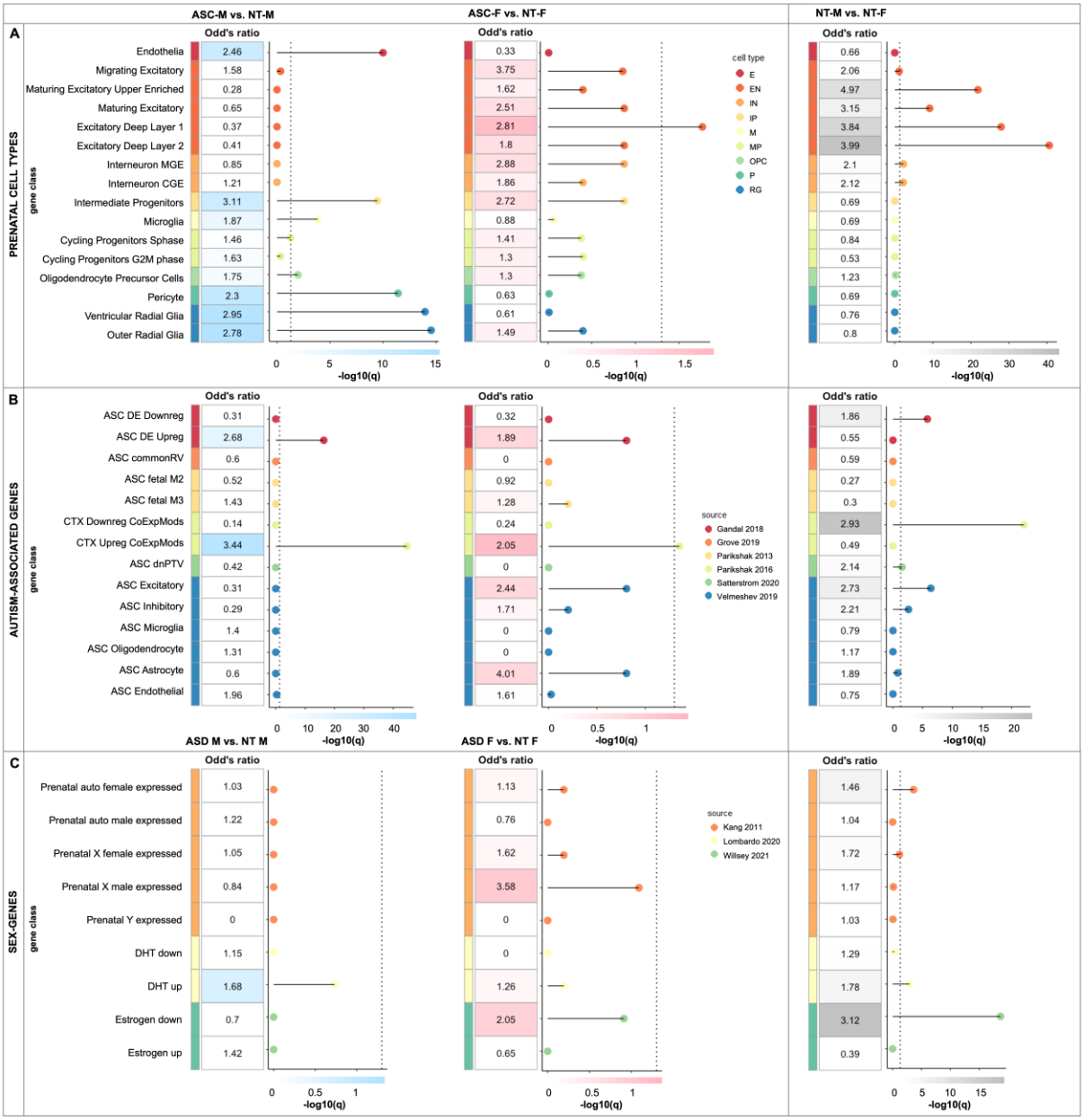
by comparing sex prediction accuracy results when only using the most salient brain regions (left panel: 'ASC-sensitive mask') versus excluding the most salient brain regions in the RAP maps (right panel: 'ASC-insensitive mask'). Abbreviations: PCA=principal component analysis, NT=neurotypical, ASC=autism, F=females, M=males, RAP=region-aligned prediction method.

FIGURE S7. Comparison of highly mis- and accurately classified autistic individuals[a]



[a] The brain maps depict the different RAP (spatial representation of the sex predictions) imaging-t-maps across **A**) highly-accurately classified vs. highly mis-classified autistic females and **B**) highly-accurately classified vs. highly mis-classified autistic males. Abbreviations: ASC=autism, F=females, M=males.

FIGURE S8. Enrichment analyses when using a more conservative threshold[a]



[a] Enrichment analysis using a more conservative list of 16,906 background genes based on real estimates of genes expressed in cortical tissue (71). Odds-ratios at an FDR-corrected $p<0.05$ resulting from the gene set enrichment analyses of RAP-imaging-t-maps (ASC-M vs. NT-M; ASC-F vs. NT-F; NT-M vs. NT-F) and associated gene lists with different classes of genes acting prenatally and relevant in the context of autism and sexual differentiation. (**A**) genes from prenatal cell types (57) (endothelia (E), excitatory neurons (EN; migrating excitatory, maturing excitatory upper enriched, maturing excitatory, excitatory deep layer 1, excitatory deep layer 2), interneurons (IN; interneuron MGE and interneuron CGE), intermediate progenitors (IP), microglia (M), mitotic progenitors (MP; cycling progenitor S-phase and cycling progenitors G2M phase), oligodendrocyte precursors (OPC), pericyte (P), and radial glia (RG; ventricular radial glia and outer radial glia), (**B**) autism-associated genes,

including common genetic variants (ASC commonRV) (58), de novo mutations (fetal gene co-expression modules [ASC fetal M2, ASC fetal M3] (61); 102 rare, de novo protein truncating genes [ASC dnPTV]) (60) and transcriptionally dysregulated genes (differentially-expressed downregulated [ASC DE Downreg], differentially-expressed upregulated [ASD DE Upreg] (62); cortical downregulated co-expression modules [CTX Downreg CoExpMods], cortical upregulated co-expression modules [CTX Upreg CoExpMods] (63); ASC Excitatory, ASC Inhibitory, ASC Microglia, ASC Oligodendrocyte, ASC Astrocyte, ASC Endothelial (64)), and (**C**) sex-differentially expressed genes acting prenatally such as genes differentially regulated by dihydrotestosterone (DHT) (67–69), estrogen (70) and (autosomal and X-/Y-chromosome-linked) sex-differential gene array expression data from prenatal samples (65, 66). Abbreviations: NT=neurotypical, ASC=autism, F=females, M=males.

Supplemental Tables

TABLE S1. ABIDE sample characterization

Variable	ASC-M			ASC-F			NT-M			NT-F			post hoc
N	526			115			532			239			-
	Mean	Std	Range	Mean	Std	Range	Mean	Std	Range	Mean	Std	Range	-
age	13.20	5.70	5.32-55.0	13.66	7.28	5.22-54.0	13.53	6.27	5.89-56.0	13.070	6.36	5.91-47.0	ASC=NT
FIQ	105.39	17.63	41.0-149.0	104.35	16.67	66.0-146.0	112.39	12.65	71.0-148.0	113.67	13.12	80.0-149.0	ASC<NT
VIQ	106.30	18.59	42.0-180.0	105.71	16.67	70.0-145.0	113.71	13.47	67.0-147.0	114.09	14.56	83.0-156.0	ASC<NT
PIQ	104.74	17.33	37.0-149.0	102.21	18.11	53.0-148.0	108.78	13.92	62.0-147.0	109.49	13.26	79.0-145.0	ASC<NT
ADI social	19.64	5.34	0.0-30.0	19.08	6.20	0.0-30.0	-	-	-	-	-	-	M=F
ADI communication	15.73	4.48	0.0-25.0	14.90	5.05	0.0-24.0	-	-	-	-	-	-	M=F
ADI RRB	5.93	2.49	0.0-12.0	5.78	2.55	0.0-12.0	-	-	-	-	-	-	M=F
ADOS-G total	11.38	3.97	2.0-22.0	11.81	3.90	3.0-21.0	-	-	-	-	-	-	M=F
ADOS-G coommunnication	3.52	1.57	0.0-8.0	3.50	1.59	0.0-7.0	-	-	-	-	-	-	M=F
ADOS-G social	7.74	2.78	2.0-14.0	7.95	2.45	2.0-14.0	-	-	-	-	-	-	M=F
ADOS-G RRB	2.11	1.54	0.0-7.0	2.02	1.55	0.0-5.0	-	-	-	-	-	-	M=F
ADOS-2 CSS	7.04	2.06	1.0-10.0	6.69	1.72	2.0-10.0	-	-	-	-	-	-	M=F
ADOS-2 SA	9.37	3.81	1.0-20.0	8.79	3.14	4.0-18.0	-	-	-	-	-	-	M=F
ADOS-2 RRB CSS	3.10	1.82	0.0-8.0	2.79	1.46	0.0-6.0	-	-	-	-	-	-	M=F

Abbreviations: ASC=autism, NT=neurotypical, M=male, F=females, FIQ=full-scale IQ, PIQ=performance IQ, VIQ=verbal IQ, ADI=Autism Diagnostic Interview, ADOS=Autism Diagnostic Observtion Schedule, CSS=calibrated severity score, SA=social-affect, RRB=restricted, repetitive behavior

TABLE S2. ADHD sample characterization

Variable	ADHD-M			ADHD-F			NT-M			NT-F			post hoc
N	324			130			225			208			-
	Mean	Std	Range	Mean	Std	Range	Mean	Std	Range	Mean	Std	Range	-
age	13.27	3.40	7.25-21.0	13.02	3.95	7.35-26.0	12.55	2.82	7.29-22.0	12.83	3.86	7.33-24.0	ADHD=NT
FIQ	102.81	16.77	55.0-149.0	100.66	16.06	48.0-134.0	114.08	15.35	58.0-158.0	112.54	14.15	75.0-144.0	ADHD<NT
VIQ	105.09	14.78	54.0-141.0	104.25	16.22	65.0-138.0	113.06	13.59	71.0-141.0	111.49	14.27	71.0-146.0	ADHD<NT
PIQ	101.82	14.76	70.0-143.0	103.86	13.65	79.0-135.0	114.41	13.69	83.0-138.0	107.71	13.69	67.0-135.0	ADHD<NT

Abbreviations: ADHD=attention-deficti/hyper-activity disorder, NT=neurotypical, M=male, F=females, FIQ=full-scale IQ, PIQ=performance IQ, VIQ=verbal IQ

TABLE S3. Summary of acquisition parameters across sites in EU-AIMS LEAP

Site	Manufacturer	Model	Software Version	Acquisition sequence	Coverage	Slices	Thickness [mm]	Resolution [mm^3]	TR [s]	TE [ms]	FA [°]	FOV
Cambridge	Siemens	Verio	Syngo MR B17	Tfl3d1_ns	256*256	176	1.2	1.1*1.1*1.2	2.3	2.95	9	270
London	GE Medical systems	Discovery mr750	LX MR DV23.1_V02_1317.c	SAG ADNI GO ACC SPGR	256*256	196	1.2	1.1*1.1*1.2	7.31	3.02	11	270
Mannheim	Siemens	TimTrio	Syngo MR B17	MPRAGE ADNI	256*256	176	1.2	1.1*1.1*1.2	2.3	2.93	9	270
Nijmegen	Siemens	Skyra	Syngo MRD13	Tfl3d1_16ns	256*256	176	1.2	1.1*1.1*1.2	2.3	2.93	9	270
Rome	GE Medical systems	Signa HDxt	24/LX/MR HD16.0_V02_1131.a	SAG ADNI GO ACC SPGR	256*256	172	1.2	1.1*1.1*1.2	5.96	1.76	11	270
Utrecht	Philips Medical Systems	Achieva/ Ingenia CX	3.2.3, 3.2.3.1	ADNI GO 2	256*256	170	1.2	1.1*1.1*1.2	6.76	3.1	9	270

TABLE S4. Summary of sample sizes employed at different stages of the model application

Sample	**Dataset**	**NT-F**	**NT-M**	**ASC-F**	**ASC-M**	**Total**
Train	ABIDE	141	141	0	0	282
Test	ABIDE	84	84	84	84	336
Test	LEAP	98	98	98	98	392
Test (auto age-matching)	LEAP	85	85	85	85	340
Sample	**Dataset**	**NT-F**	**NT-M**	**ADHD-F**	**ADHD-M**	**Total**
Test (manual age-matching)	ADHD	130	130	130	130	520
Test (auto age-matching)	ADHD	125	125	125	125	500

Abbreviations: ASC=autism, NT=neurotypical, M=male, F=females, ADHD=attention-deficti/hyper-activity disorder

TABLE S5. Sex-specific prediction accuracies in ADHD

Cohort		**ADHD200 + NeuroImage**		
Group		NT	ADHD-F	ADHD-M
A – Manual age-matching *before* the model application				
Sex-specific accuracy	Mean	0.733	0.722	0.730
	SD	0.046	0.069	0.064
B – Auto-age-matching *within* the model application				
Sex-specific accuracy	Mean	0.730	0.722	0.712
	SD	0.045	0.066	0.062

Abbreviations: ADHD=attention-deficti/hyper-activity disorder, NT=neurotypical, M=male, F=females

TABLE S6. Comparison of autistic individuals with and without ADHD

Group		NT	ASC-F		ASC-M	
Sub-group		**-**	**ADHD+**	**ADHD–**	**ADHD+**	**ADHD–**
Sex-specific accuracy	Mean	0.799	0.714	0.747	0.866	0.863
	SD	0.045	0.11	0.109	0.092	0.086

Abbreviations: ASC=autism, NT=neurotypical, M=male, F=females, ADHD=attention-deficti/hyper-activity disorder, ADHD+= autistic individuals with ADHD, ADHD-=autistic indivduals without ADHD

TABLE S7. Sex-specific prediction accuracies across control analyses

Group		NT	ASC-F	ASC-M
A – Auto-age-matching				
Sex-specific accuracy	Mean	0.800	0.737	0.863
	SD	0.024	0.053	0.041
B – Auto-age- + brain volume-matching				
Sex-specific accuracy	Mean	0.734	0.655	0.843
	SD	0.050	0.100	0.062
C – Principal component analysis and logistic regression				
Sex-specific accuracy	Mean	0.647	0.568	0.686
	SD	0.037	0.049	0.052
D – RAP verification				
ASC-sensitive mask	Mean	0.701	0.620	0.763
sex-specific accuracy	SD	0.030	0.056	0.056
ASC-insensitive mask	Mean	0.66	0.638	0.702
sex-specific accuracy	SD	0.037	0.060	0.063

Abbreviations: ASC=autism, NT=neurotypical, M=male, F=females, RAP=region-aligned prediction

TABLE S8. Most differentiating regions between NT-M and NT-F

Grey Matter region	hemisphere	size	mean	std
Temporal Fusiform Cortex, posterior division	B	22748	0.674848	0.155339
Planum Temporale	L	12209	0.667778	0.121297
Caudate	L	3949	0.602529	0.136994
Heschl's Gyrus (includes H1 and H2)	L	6084	0.676093	0.13534
Parietal Operculum Cortex	B	12782	0.661188	0.118787
Parahippocampal Gyrus, posterior division	B	14834	0.594444	0.130176
Hippocampus	R	6165	0.595922	0.090629
Supramarginal Gyrus, posterior division	B	32114	0.611748	0.10221
Cerebellum Crus I	L	26823	0.586231	0.108581
Cerebellum Vermis Crus I	B	5	0.59817	0.006328
Cerebellum Vermis VI	B	3736	0.581022	0.055262
Temporal Occipital Fusiform Cortex	B	19047	0.58575	0.125005
Superior Temporal Gyrus, posterior division	B	17277	0.610472	0.100785
Angular Gyrus	B	29354	0.580459	0.095334
Planum Polare	B	9013	0.565302	0.094504
Inferior Temporal Gyrus, posterior division	B	32133	0.559189	0.118362
Accumbens	L	756	0.551552	0.067611
Supramarginal Gyrus, anterior division	B	22159	0.590058	0.130507
Cerebellum I-IV	R	9228	0.558872	0.109159
Central Opercular Cortex	B	19707	0.530095	0.160716

White Matter connection	hemisphere	size	mean	std
Sagittal stratum (include inferior longitidinal fasciculus and inferior fronto-occipital fasciculus)	R	2228	0.744193	0.07272
Sagittal stratum (include inferior longitidinal fasciculus and inferior fronto-occipital fasciculus)	L	2231	0.694	0.072734
Tapetum	L	600	0.666728	0.032677
Retrolenticular part of internal capsule	R	2515	0.636811	0.095918
Medial lemniscus	R	690	0.662763	0.049701
Tapetum	R	596	0.636833	0.04043
Cingulum (hippocampus)	R	1236	0.60971	0.1172
Retrolenticular part of internal capsule	L	2469	0.569462	0.114106
Posterior corona radiata	R	3728	0.591619	0.065996
Corticospinal tract	R	1362	0.580639	0.067095
Fornix (cres) / Stria terminalis (can not be resolved with current resolution)	R	1124	0.574191	0.062119
Anterior limb of internal capsule	L	3018	0.558917	0.085043
Cingulum (hippocampus)	L	1155	0.564226	0.071553
Genu of corpus callosum	B	8851	0.542123	0.104349
Posterior corona radiata	L	3714	0.547375	0.072429
Splenium of corpus callosum	B	12729	0.539905	0.056821
Cerebral peduncle	R	2278	0.530177	0.063799
Inferior cerebellar peduncle	R	968	0.536343	0.060001
Posterior thalamic radiation (include optic radiation)	L	3978	0.515662	0.16168

Abbreviations: NT=neurotypical, M=male, F=female, R=right, L=left, B=both

TABLE S9. Most male-shifted regions in ASC-F

Grey Matter region	hemisphere	size	mean	std
Cerebellum Crus I	L	26823	0.255442	0.072154
Heschl's Gyrus (includes H1 and H2)	B	6084	0.226662	0.104109
Caudate	L	3949	0.188232	0.110903
Planum Temporale	L	12209	0.183346	0.134065
Occipital Fusiform Gyrus	B	28100	0.204175	0.133693
Amygdala	L	2662	0.168623	0.063565
Parietal Operculum Cortex	B	12782	0.171952	0.086195
Hippocampus	L	6154	0.165365	0.0567
Accumbens	L	756	0.171228	0.040709
Inferior Temporal Gyrus, temporooccipital part	B	20013	0.157296	0.083235
Cerebellum VI	L	17861	0.172028	0.094903
Inferior Temporal Gyrus, posterior division	B	32133	0.146074	0.114158
Caudate	R	4127	0.142831	0.092932
Cerebellum Crus II	L	21227	0.141174	0.089322
Central Opercular Cortex	B	19707	0.132469	0.113722
Temporal Occipital Fusiform Cortex	B	19047	0.137184	0.08119
Lateral Occipital Cortex, inferior division	B	57896	0.139874	0.107609
Superior Temporal Gyrus, posterior division	B	17277	0.122037	0.156537
Planum Polare	B	9013	0.132844	0.079657
Parahippocampal Gyrus, posterior division	B	14834	0.116739	0.086311

White Matter connection	hemisphere	size	mean	std
Genu of corpus callosum	B	8851	0.272063	0.102349
Retrolenticular part of internal capsule	R	2515	0.27698	0.06932
Sagittal stratum (include inferior longitidinal fasciculus and inferior fronto-occipital fasciculus)	R	2228	0.221877	0.115045
Tapetum	L	600	0.203539	0.110326
Retrolenticular part of internal capsule	L	2469	0.218705	0.063159
Fornix (cres) / Stria terminalis (can not be resolved with current resolution)	L	1125	0.207553	0.044443
Sagittal stratum (include inferior longitidinal fasciculus and inferior fronto-occipital fasciculus)	L	2231	0.195957	0.039052
Tapetum	R	596	0.199917	0.020729
Fornix (cres) / Stria terminalis (can not be resolved with current resolution)	R	1124	0.17257	0.101445
Uncinate fasciculus	L	376	0.164049	0.029604
Superior longitudinal fasciculus	R	6607	0.167562	0.059507
Superior cerebellar peduncle	R	992	0.137128	0.092553
Posterior thalamic radiation (include optic radiation)	L	3978	0.119328	0.145831
Cingulum (hippocampus)	L	1155	0.130622	0.023392
Posterior corona radiata	R	3728	0.13005	0.034196
Posterior thalamic radiation (include optic radiation)	R	3972	0.138363	0.095073
Uncinate fasciculus	R	380	0.120574	0.022961
Medial lemniscus	R	690	0.137374	0.06416
Corticospinal tract	R	1362	0.132608	0.080538

Abbreviations: ASC=autism, F=female, R=right, L=left, B=both

TABLE S10. Most male-shifted regions in ASC-M

Grey Matter region	hemisphere	size	mean	std
Amygdala	R	3215	0.17297	0.054697
Pallidum	R	2118	0.164403	0.02943
Accumbens	R	666	0.159551	0.022322
Thalamus	L	11488	0.158337	0.033188
Cerebellum VIIIa	R	9826	0.137376	0.070234
Cingulate Gyrus, anterior division	B	31689	0.149518	0.04923
Thalamus	R	11186	0.14557	0.035606
Putamen	R	6397	0.132497	0.046621
Caudate	R	4127	0.130855	0.027515
Subcallosal Cortex	B	16466	0.131314	0.047305
Cerebellum VIIb	R	9885	0.115508	0.065721
Frontal Medial Cortex	B	12100	0.112058	0.070862
Cerebellum VIIIb	R	8185	0.114916	0.055648
Juxtapositional Lobule Cortex (formerly Supplementary Motor Cortex)	B	17545	0.113293	0.042752
Pallidum	L	2133	0.105007	0.049254
Cingulate Gyrus, posterior division	B	35642	0.100388	0.051582
Temporal Pole	B	62256	0.095262	0.061998
Accumbens	L	756	0.097188	0.027359
Amygdala	L	2662	0.0988	0.055703
Paracingulate Gyrus	B	31069	0.096296	0.052689

White Matter connection	hemisphere	size	mean	std
Superior corona radiata	R	7500	0.195155	0.033281
Superior fronto-occipital fasciculus (could be a part of anterior internal capsule)	R	507	0.191542	0.009216
Anterior corona radiata	R	6849	0.172089	0.041197
Cerebral peduncle	L	2278	0.166158	0.034835
Cingulum (cingulate gyrus)	R	2342	0.168892	0.041982
Posterior limb of internal capsule	R	3754	0.154056	0.041451
Genu of corpus callosum	B	8851	0.143966	0.062544
Cerebral peduncle	R	2278	0.140164	0.082683
Body of corpus callosum	B	13711	0.143612	0.033902
Posterior limb of internal capsule	L	3752	0.137433	0.026209
Superior longitudinal fasciculus	R	6607	0.128455	0.082359
Anterior limb of internal capsule	R	3138	0.14209	0.02755
Fornix (column and body of fornix)	B	659	0.128445	0.031074
Retrolenticular part of internal capsule	L	2469	0.122443	0.055491
Fornix (cres) / Stria terminalis (can not be resolved with current resolution)	L	1125	0.121074	0.058493
Superior fronto-occipital fasciculus (could be a part of anterior internal capsule)	L	507	0.121725	0.033861
Superior corona radiata	L	7508	0.111048	0.041673
Cingulum (cingulate gyrus)	L	2751	0.112687	0.035577

Abbreviations: ASC=autism, M=male, R=right, L=left, B=both